

Effect of Proline and its Derivatives on the Properties of Silk Fibroin Microneedles

Bin Tian, Yu-Min Zhang, Zhen-Zhen Qi, Le-Hao Zhang,
Zu-Qiang Yin, Shen-Zhou Lu*

*National Engineering Laboratory for Modern Silk, College of Textile and Clothing Engineering,
Soochow University, 215127, Suzhou, China*

Abstract

With its good biocompatibility and excellent mechanical properties, silk fibroin microneedles can transport drugs to the body fluid circulation system and then act on the affected area, so as to replace intravenous injection and oral administration, and achieve the purpose of treating diseases. In the process of processing and use, silk fibroin microneedles are non-toxic, harmless, pollution-free and biodegradable to human body and environment. Therefore, the application prospect and application range of silk fibroin microneedles are very wide.

In this paper, the effects of proline and its derivatives prolinamide and hydroxyproline on the performance of silk fibroin microneedles were studied on the basis of the previous experiments of constructing microneedles to carry drugs. The composite silk fibroin microneedles were obtained by pouring the amino acid/silk fibroin mass ratio of 0/10, 1/10, 2/10, 3/10 and 4/10 into a polydimethylsiloxane mold, after vacuum defoaming and drying. The length of the microneedles was about 600 μm . The aggregation structure of amino acid/silk fibroin microneedles was measured by X-ray diffraction (XRD), Fourier transform infrared spectroscopy (FTIR) and Raman Scattering Spectroscopy. The mechanical properties of the microneedles were measured by texture analyzer. The results showed that: (1) The silk fibroin microneedles prepared by adding proline and its derivatives had predominately Silk I crystal structure; (2) When the mass ratio of proline and its derivatives to silk fibroin reached 2/10, it had a higher swelling degree and a lower dissolution rate; (3) The silk fibroin microneedles prepared by proline and its derivatives have good mechanical properties. The following conclusion was drawn: with the addition of proline and its derivatives, silk fibroin microneedles with higher swelling degree and lower dissolution rate can be obtained. The crystal structure of Silk I is formed inside the microneedles, which has good penetration and fracture properties. It is expected that the microneedles can be used as swelling microneedles for drug transdermal delivery.

Keywords: Silk Fibroin; Microneedle; Proline; Silk I Crystalline Structure

*Corresponding author.

Email address: lushenzhou@suda.edu.cn (Shen-Zhou Lu).

1 Introduction

In the past few decades, transdermal delivery has been an attractive route for drug delivery [1-3]. Skin is the largest organ in the human body. It receives one-third of the blood supply of the entire body and was not used as a drug delivery route until the end of the 20th century. The mechanism of microneedle transdermal drug delivery is to use microneedles to penetrate the tightly arranged stratum corneum of the skin [4], and directly deliver the drug to the dermis. Microneedle penetration strengthens the drug delivery channel into the skin, and the microneedle will not penetrate too deep, which can protect the human skin. The administration process is almost painless, which can improve the compliance of patients. In addition, the dosage of drugs delivered by skin is usually lower than oral drugs, which can avoid the side effects caused by unstable absorption and metabolism of drugs in gastrointestinal tract [5]. In the case where oral drug delivery is difficult, transdermal drug delivery technology can be easily applied on the skin, and provide effective blood concentration level [6].

Different forms of microneedles have different ways of releasing drugs. There are five types of microneedles: solid microneedles, hollow microneedles, swelling microneedles, soluble microneedles, and coated microneedles [7]. The main advantage of solid microneedles is their firmness, which makes it easier to penetrate the skin and can be used to pretreat the skin. Henry et al [8]. First demonstrated the increase of transdermal flux of calcein after the silicon microneedles were prepared by ion etching. Dissolving microneedles are mainly made of polymers or polysaccharides, which release drugs through dissolution after piercing the skin [9]. Hollow microneedles have the same empty cavity as a traditional hypodermic needle to deliver drugs to the skin or through the skin into the blood, but the microneedles are shorter and the flow rate can be controlled by a micropump or syringe [10]. Coated microneedles are made from a substrate coated with drugs that can be delivered quickly to the skin and increase the long-term stability of the active drug, although the amount of drug can be uncontrolled. Swelling microneedles can absorb the interstitial fluid of the skin and provide channels for drug delivery in the microneedles. When used, the integrity can be retained without residual accumulation in the skin, and it is convenient for patients to use and improve patient dependence. Swelling microneedles have the function of rate control and prolong the time of drug administration by adjusting the swelling performance [11]. Now, the most widely used is the use of microneedles to carry drugs, after the microneedle enters the epidermis, the needle body swells and the drugs enter the human skin.

Silk fibroin protein is a kind of natural polymer material with good biocompatibility and natural degradation, and rarely has sensitization reaction [12]. Silk fibroin protein can also maintain the biological activity of the drugs it carries for a long time, so it is extremely suitable for the preparation of biological materials [13]. Based on previous studies by scholars, it can be known that pure silk fibroin protein microneedles have good mechanical properties and can successfully achieve the purpose of drug release by microneedles. However, the untreated pure silk fibroin is random coil structure, its molecular chain arrangement is disordered, and water molecules can enter smoothly, which makes the microneedles dissolve quickly when contacting body fluid. In this paper, we explored the effects of proline and its derivatives on the structure and properties of silk fibroin microneedles, and explored a more balanced addition strategy to prepare the swelling silk fibroin microneedles with low solubility, high swelling and good mechanical properties. It provides new methods and ideas for the improvement of microneedle transdermal drug delivery materials.

75 **2 Materials and Methods**

76 **2.1 Experimental Materials and Instruments**

77 **2.1.1 Experimental Materials**

78 Silkworm cocoon shell (Suzhou siruibao Biotechnology Co., Ltd., China), sodium carbonate,
79 sodium bicarbonate, lithium bromide (Tiancheng Chemical Co., Ltd., China), dialysis bag
80 (MWCO: 8-12kd, Pierce), L-proline (Shanghai Yuanye Biotechnology Co., Ltd.), L-prolinamide
81 (Shanghai Aladdin Biochemical Technology Co., Ltd.), L-hydroxyproline (Shanghai Aladdin Bio-
82 chemical Technology Co., Ltd.) etc.

83 **2.1.2 Experimental Instruments**

84 FA2004 electronic balance, SHY-2 digital display water bath constant temperature oscillator,
85 DT5-2 low speed table centrifuge, DHG-9246A electrothermal constant temperature blast drying
86 oven, 84-1A magnetic stirrer, SmartSpec Plus spectrophotometer, X'PERT-PRO MPD X-ray
87 diffractometer, Nicolet is 5 intelligent Fourier transform infrared spectrometer, TMS-PRO texture
88 analyzer.

89 **2.2 Experimental Part**

90 **2.2.1 Preparation of Silk Fibroin Solution**

91 Weigh a certain proportion of sodium carbonate and sodium bicarbonate and dissolve in water at
92 100 °C. Add 80 g cocoon shell and cook for 30 min. Clean boiled cocoons with deionized water to
93 remove sericin from the surface. The cleaned cocoon shell is boiled in sodium carbonate/sodium
94 bicarbonate buffer solution again for 30 min and cleaned. Repeat three times. Put the degummed
95 silk fibroin into 60 °C oven for drying. Weigh 15 g of dried degummed silk fibroin and dissolve it
96 in LiBr solution (9.3 M) at 65 °C for 60 min. After cooling, the solution was put into a dialysis
97 bag and placed in 4 °C deionized water for dialysis for 3-4 days. The obtained silk fibroin solution
98 was stored in a 4 °C refrigerator for later use.

99 **2.2.2 Preparation of Microneedles by Die Casting**

100 Proline and its derivatives were prepared into an aqueous solution with a concentration of 150
101 mg/mL for use. Mix according to the mass ratio of amino acids to silk fibroin of 0/10, 1/10,
102 2/10, 3/10, 4/10, stir fully, and pour the mixed solution into the PDMS mold. After vacuuming
103 for several times, the formed microneedles were dried for 24 h in a constant temperature and
104 humidity room.

105 **2.2.3 X-ray Diffraction**

106 X-ray diffraction sample preparation: The silk fibroin fibers were cut into fine powder particles
107 and various silk fibroin microneedles were cut into pieces with scissors and screened by 80 μm
108 sieve. The particles that could pass the sieve were used for testing.

109 X-ray diffraction analysis: Using automatic X'PERT PRO MPD X-ray diffractometer. The
110 diffraction intensity curve was recorded between 5° and 45° under the conditions of 10°/min
111 scanning speed, 40 kV, and 30 mA.

112 2.2.4 Infrared Detection

113 The prepared silk fibroin film was tested on the Nicolet is 5 intelligent Fourier transform infrared
114 spectrometer. The scanning range was 400~4000 cm⁻¹, and the infrared absorption spectrum
115 was obtained.

116 2.2.5 Raman Scattering Spectroscopy

117 Raman spectrum was measured using a Japanese HORIBA Raman Microscopy (HORIBA Ltd.,
118 Kyoto City, Japan). The excitation wavelength was 532 nm, slit width was 100 μm, and 1200
119 gr/mm grating was selected. The scanning time of the fixed sample was 20 s, and the Raman
120 scattering spectrum recording step range was 200~2000 cm⁻¹.

121 2.2.6 Moisture content of microneedles

Different amino acids/silk fibroin protein microneedles were weighed, denoted as M1, placed in
an oven at 105 °C, dried for 2 hours, and weighed M2. The moisture content of the microneedle
was calculated by the formula (1).

$$\text{Moisture content} = \frac{M1 - M2}{M2} * 100\% \quad (1)$$

122 2.2.7 Detection of Dissolution and Swelling of Microneedles

The silk fibroin microneedle was balanced at the same room temperature for 24 hours, then the
M1 was weighed and put into a centrifuge tube. PBS buffer (pH = 7.4) was added at a bath ratio
of 1:100(W/V), and the silk fibroin microneedle was shaken in a constant temperature shaker
at 37 ° water bath for 24 hours. Clean the solid silk fibroin microneedles in the solution with
deionized water for 3 times, then absorb surface moisture with absorbent paper and weigh M2.
After the solution in the centrifuge tube was centrifuged at 3000 r/min, the supernatant was taken
and the absorbance of the solution at 278 nm was measured by ultraviolet spectrophotometer.
The swelling rate Q(%) and silk fibroin dissolution rate C(%) of microneedles were calculated by
formula (2) and (3) respectively.

$$Q = \frac{M2 - M1 \times (1 - F)}{M1 \times (1 - F)} \times 100\% \quad (2)$$

$$C = \frac{KAV}{M1 \times (1 - F)} \times 100 \quad (3)$$

123 Q—Silk fibroin swelling degree (%);

124 M1—The initial weight of the sample (g);

125 M2—Weight of sample after soaking (g);

- 126 F—Moisture content (%);
 127 C—The dissolution rate of c-silk fibroin (%);
 128 K—UV absorption constant of silk fibroin solution;
 129 A—Absorbance;
 130 V—Volume of PBS solution (mL).

131 2.2.8 Measurement of Compression Strength of Microneedles

132 Different amino acid/silk fibroin microneedles were evenly cut into 3×3 array, 5 parallel samples
 133 in each group, the needle tip was placed upward under the TMS-PRO texture analyzer, and
 134 the breaking strength of the needle tip was detected. Test conditions: initiation force 0.02 N,
 135 maximum detection range 25 N, deformation 80%, compression speed 10 mm/min.

136 3 Experimental Results and Analysis

137 3.1 Silk Fibroin Protein Blend Membrane

138 In order to clearly observe whether proline and its derivatives are compatible with silk fibroin,
 139 silk fibroin blend membrane was prepared.

140 As shown in Fig. 1, proline and prolinamide are better fused with silk fibroin in any proportion.
 141 When the ratio of hydroxyproline to silk fibroin reached 3/10, white precipitates appeared on the
 142 film, which was the self-crystallization of small molecules. This indicates that when the proportion
 143 is high, the compatibility between hydroxyproline and silk fibroin becomes poor, and this kind
 144 of phase separation material is not suitable for microneedle material because of its unstable
 145 structure.

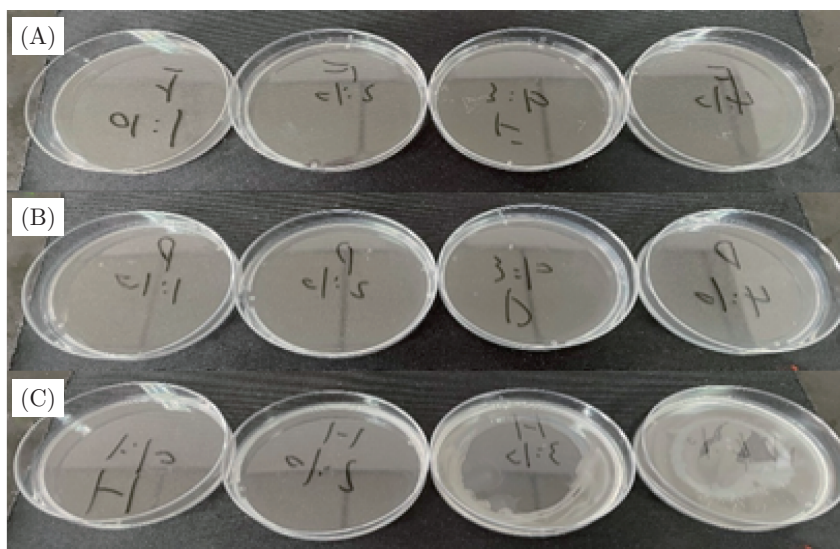


Fig. 1: Silk fibroin protein films of different amino acids: (A) proline/silk fibroin, (B) prolinamide/silk fibroin, (C) hydroxyproline/silk fibroin

146 3.2 Morphology of Microneedles

147 The microneedle mold determines the external shape of the microneedle. Here, we used the
148 mold of the same specification to make the microneedle prepared by adding different amino acid
149 silk protein solutions have the same needle shape. As shown in Fig. 2(A), the microneedle patch
150 consists of 225 (15×15) microneedles evenly distributed. In Fig. 2(B), we can see the microneedles
151 prepared by the mold were slender and tapered, with a length of about $600 \mu\text{m}$.

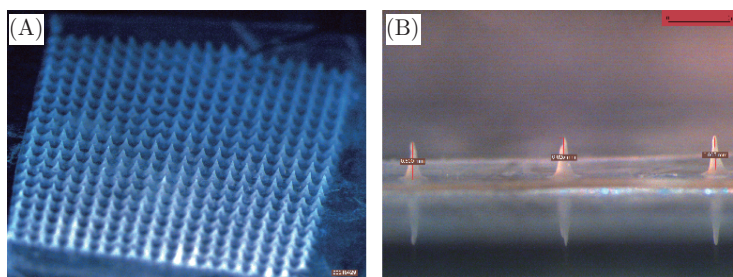


Fig. 2: Morphology of microneedles

152 3.3 X-ray Diffraction Analysis

153 In the X-ray diffraction pattern, the crystal peaks of Silk I appeared at 12.2° , 19.7° , 24.7° , 28.2° ,
154 32.3° , 36.8° and 40.1° , while the crystal peaks of Silk II appeared at 9.1° , 18.9° , 20.7° and 24.3°
155 [14].

156 As shown in Fig. 3(A), the X-ray diffraction curve of pure silk fibroin microneedles shows a
157 steamed bread shape without obvious absorption peak. The results showed that the molecular

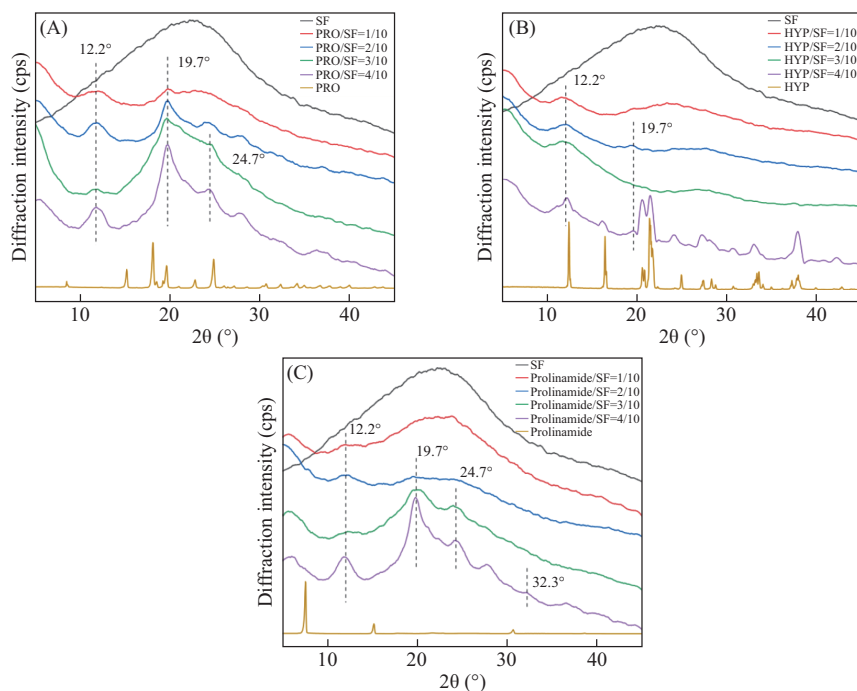


Fig. 3: X-ray diffraction curves of amino acid/silk fibroin microneedles: (A) proline/silk fibroin, (B) hydroxyproline/silk fibroin, (C) prolinamide/silk fibroin

158 chain of silk fibroin was in random coil state, and no crystal was formed in the microneedles.
159 Therefore, pure silk fibroin microneedles are easily soluble in water. When the ratio of proline
160 to silk fibroin is 1/10, 2/10, 3/10 and 4/10, the crystal peaks appear at 12.2° and 19.7°. With
161 the increase of the proportion of proline, the peak pattern becomes more acute, indicating the
162 formation of the crystal structure of Silk I. Proline mainly induces silk fibroin protein to form
163 the crystal structure of Silk I. The addition of proline greatly reduced the dissolution rate of silk
164 fibroin microneedles.

165 In Fig. 3(B), when the ratio of hydroxyproline to silk fibroin was 1/10, the curve showed
166 no significant difference from that of pure silk fibroin, presenting a state of random coiling. At
167 2/10, 3/10 and 4/10, the crystal peaks of 12.2° and 19.7°. As the proportion of hydroxyproline
168 increases, the peak shape becomes sharper. When the ratio of hydroxyproline to silk fibroin was
169 4/10, a large number of crystallization peaks of hydroxyproline appeared. The above results
170 indicate that when hydroxyproline/silk fibroin is at 1/10, hydroxyproline does not induce silk
171 fibroin to form Silk I crystallization. When the ratio reached 2/10, 3/10 and 4/10, the silk fibroin
172 protein mainly formed the crystal structure of Silk I under the induction of hydroxyproline. When
173 hydroxyproline/silk fibroin is 4/10, hydroxyproline begins to crystallize, which indicates that the
174 compatibility between hydroxyproline and silk fibroin protein is poor at this time. This is the
175 same as with the blending film.

176 In Fig. 3(C), 1/10 prolinamide/silk fibroin barely changed compared to pure silk fibroin. At
177 2/10, 3/10 and 4/10, the crystal peaks of Silk I appeared at 12.2° and 19.7°, and gradually became
178 sharp with the increase of prolinamide dosage. When reaching 3/10, the crystal peaks of 24.7°
179 and 32.3° appeared, indicating that a large number of crystal structures of Silk I were formed
180 at this time. It can be seen from the figure that prolinamide has good compatibility with silk
181 fibroin, and induces silk fibroin to mainly form the crystal structure of Silk I.

182 3.4 Infrared Detection

183 In order to further explore the aggregation structure of the modified silk fibroin microneedle,
184 Fourier transform infrared spectrometer was used to test its secondary structure. In the infrared
185 spectrum, the untreated silk fibroin microneedle had obvious absorption peaks at 1635 cm⁻¹, 1530
186 cm⁻¹ and 1235 cm⁻¹, which were characteristic peaks of random coil.

187 In the infrared spectrum, it can be seen that the silk fibroin microneedles have no other obvi-
188 ous absorption peaks compared with pure silk fibroin microneedles after adding proline and its
189 derivatives. Absorption peaks were observed at 1635 cm⁻¹ (amide I), 1515 cm⁻¹ (amide II) and
190 1235 cm⁻¹ (amide III) [15]. With the increase of amino acids, the absorption peaks of silk fibroin
191 in the amide I, the amide prism and the amide prism remain basically unchanged. In general, it
192 is difficult to distinguish random coil from α -helix directly by FTIR. By combining FTIR spec-
193 tra with XRD data, we can determine that the absorption peak should be the coexistence and
194 superposition of α -helix, random coil and β -folding peak.

195 3.5 Raman Scattering Spectroscopy

196 To further illustrate the effect of proline and its derivatives on silk fibroin structure, we examined
197 the changes of Raman spectra of silk fibroin material.

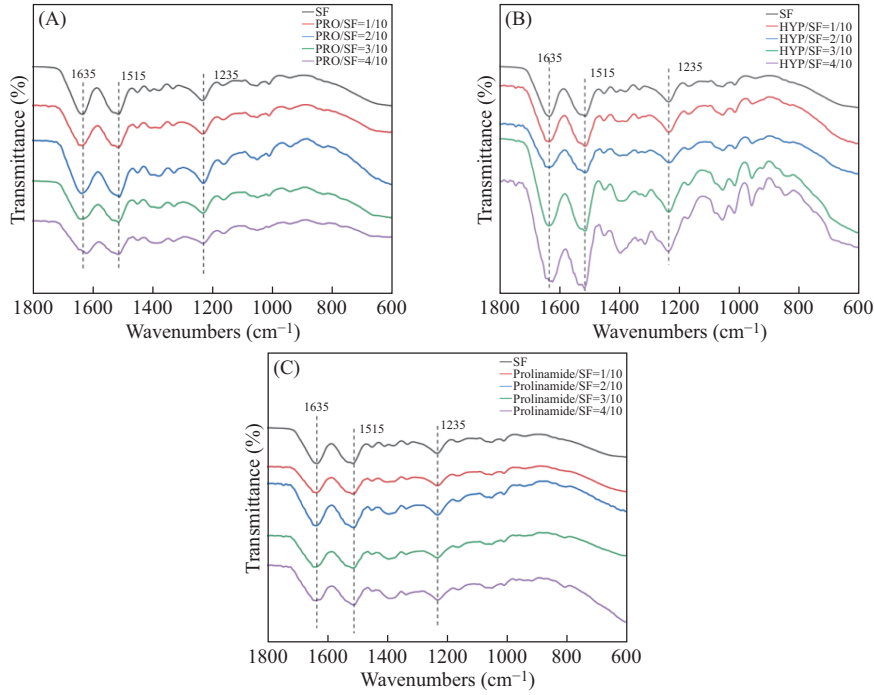


Fig. 4: FTIR of proline and its derivatives/silk fibroin microneedles: (A) proline/silk fibroin, (B) hydroxyproline/silk fibroin, (C) prolinamide/silk fibroin

198 In Raman spectroscopy (Fig. 5), it can be seen that proline and its derivatives have scattering
 199 peaks at 1667 cm^{-1} (amide I), 1245 cm^{-1} (amide III) and 1106 cm^{-1} [16]. With the increasing
 200 proportion of amino acids, the absorption peaks of silk fibroin at amide I and amide III were

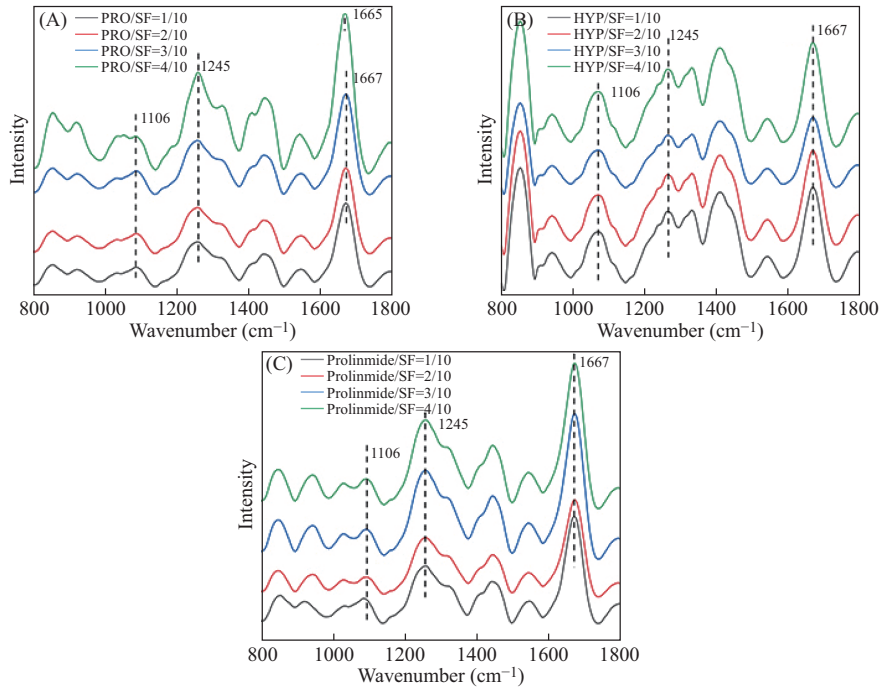


Fig. 5: Raman spectroscopy of proline and its derivatives/silk fibroin microneedles: (A) proline/silk fibroin, (B) hydroxyproline/silk fibroin, (C) prolinamide/silk fibroin

201 basically unchanged, which was a typical random curly conformation. Combining XRD data and
 202 FIRT spectra, we can further confirm that the structure of silk fibroin protein consists of a large
 203 amount of α -helix, random curl and a small amount of β -fold after the addition of proline and its
 204 derivatives.

205 3.6 Moisture content of microneedles

206 Tukey test was used for moisture content of different proportions of amino acids. As shown in Fig.
 207 6(A), it can be found that the moisture content of different proportions of proline is about 9%. As
 208 shown in Fig. 6(B), the water content of hydroxyproline varies greatly with different proportions,
 209 and when the ratio of amino acids to silk fibroin reaches 3/10, the water content is 6.5%, which is
 210 the minimum. When reaching 4/10, the moisture content is the largest, which can reach 10.6%.
 211 In Fig. 6(C), it can be seen that there are significant differences between prolinamide. At 2/10,
 212 the moisture content is 14.7%, while at 1/10, the moisture content is only 6%.

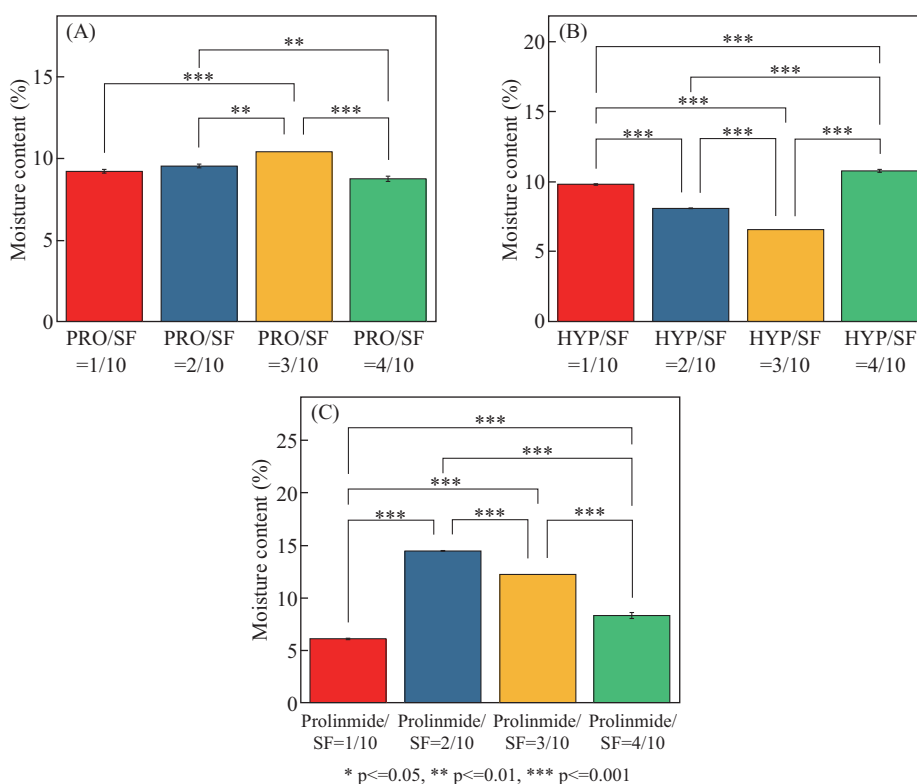


Fig. 6: Moisture content of microneedles: (A) proline/silk fibroin, (B) hydroxyproline/silk fibroin, (C) prolinamide/silk fibroin

213 3.7 Swelling and Dissolution Properties of Microneedles

214 Proline and its derivatives were added and mixed with silk fibroin protein to obtain silk fibroin
 215 microneedle with high swelling degree. By controlling the ratio of drugs and silk fibroin, different
 216 microneedles were prepared to compare their swelling and dissolution properties. In order to
 217 simulate the internal environment of human body, we soaked microneedles in PBS solution (pH
 218 = 7.4). Fig. 7 and Fig. 8 use Tukey test for significant difference analysis.

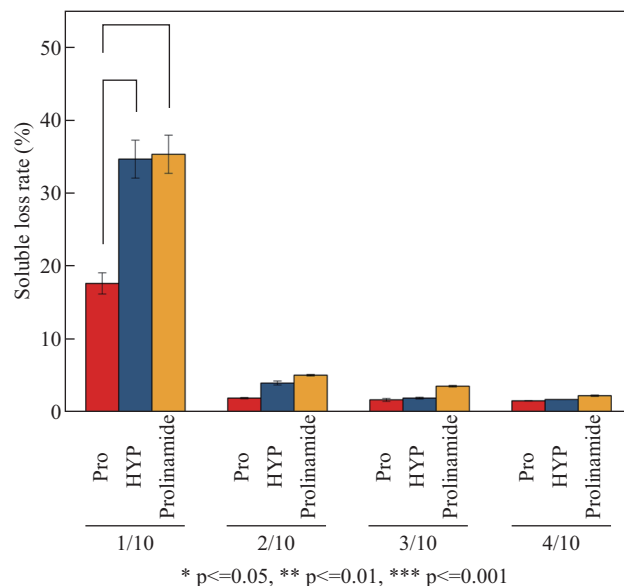


Fig. 7: Dissolution property of silk fibroin microneedles

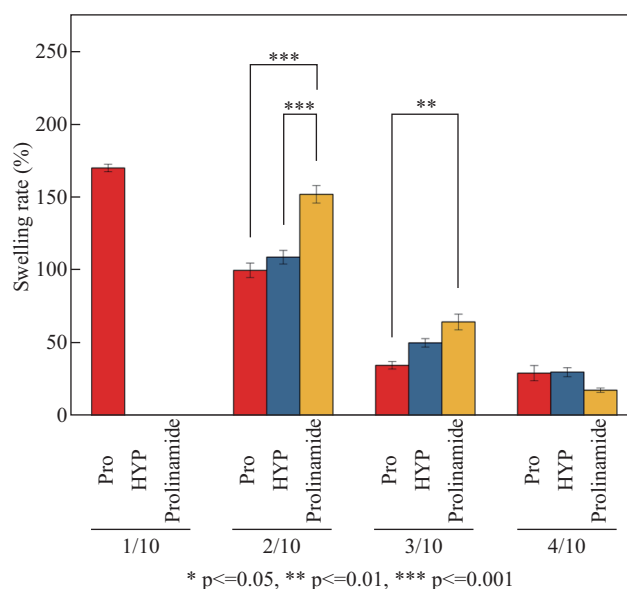


Fig. 8: Swelling properties of silk fibroin microneedles

219 It can be seen from Fig. 7 that when the ratio of proline and its derivatives to silk fibroin is
 220 2/10, 3/10 and 4/10, the dissolution loss of the three microneedles is very small, less than 6%, and
 221 there is no significant difference. But at 1/10, hydroxyproline and prolinamide were significantly
 222 different from proline. At this time, the dissolution rate of hydroxyproline and prolinamide is as
 223 high as 35%, while that of proline is only 18%. This is because when hydroxyproline/silk fibroin
 224 and prolinamide/silk fibroin are 1/10, their structures are basically random coil, they are easily
 225 soluble in water, and the loss of protein is large, so the dissolution rate is relatively large.

226 Pure silk fibroin microneedles dissolve rapidly in PBS solution, so it is impossible to measure its
 227 swelling degree. When the ratio of hydroxyproline/silk fibroin and prolinamide/silk fibroin was
 228 1/10, the silk fibroin microneedles dissolved rapidly in solution, so the swelling degree could not

229 be measured. It can be seen from Fig. 8 that when proline and its derivatives/silk fibroin is 4/10,
230 the swelling of the three microneedles is about 25%, and there is no significant difference. In the
231 case of no significant difference in dissolution rate, there was a significant difference in swelling
232 rate when amino acid/silk fibroin was 2/10. When the amino acid/silk fibroin ratio was 2/10, the
233 swelling degree of silk fibroin microneedle was up to 150% when prolinamide was added, while
234 the swelling degree of other microneedles were only about 100%. It can be considered to use the
235 additive amount of 2/10 to get the higher swelling rate of silk fibroin microneedles. Prolinamide
236 microneedles at 2/10 were significantly different from other microneedles. In addition, when the
237 ratio of amino acid to silk fibroin was 3/10, the swelling degree of microneedles with prolinamide
238 was 60%, while that of proline was only 34%.

239 With the increase of amino acid content, the swelling and dissolution of micro needle decreased
240 gradually. This is because the small molecules of proline, hydroxyproline and prolinamide promote
241 the formation of a small amount of silk fibroin crystallization, thus reducing the loss rate of silk
242 fibroin, so it has certain swelling and insoluble. When PBS solution is added, the non-crystalline
243 region expands, and a small amount of crystal region suppresses its infinite expansion. With the
244 increase of the small amino acid molecules, silk fibroin was induced to produce more crystallized
245 regions. The results showed that the dissolution rate of microneedle decreased with the increase
246 of amino acid small molecule. With the increase of addition dose, hydroxyl, peptide bond and
247 amino group in hydroxyproline, proline and prolinamide molecules can interact with polar groups
248 such as amide bond, Unbound hydroxyl group or carboxyl group in silk fibroin molecular chain
249 to form hydrogen bond. The formation of these hydrogen bonds further increases the interaction
250 of silk fibroin proteins, so the swelling properties of microneedles also tend to decrease.

251 3.8 Mechanical Properties of Microneedles

252 Microneedles must be strong enough to penetrate the cuticle of the skin for their therapeutic
253 purpose. The needle must not break in order to deliver the drug to the dermis, where it can
254 enter the body's fluid circulation for transdermal release. Fig. 9 shows the shape changes of
255 the microneedle before and after the strength test. When testing the mechanical properties of
256 the microneedle with texture instrument, the sensor will sense two forces in the process from the
257 contact of the sensor to the complete bending of the microneedle: the first is the bending force
258 of the microneedle tip, and the second is the force of the whole microneedle body being broken.
259 Here, the texture instrument records the first force, namely the breaking strength of the tip, to
260 investigate the penetration performance of the microneedle, as can be seen in Fig. 10.

261 As shown in Fig. 9(A), the 3*3 microneedle array is not compressed. The microneedles in Fig.
262 9(A) have full tips and similar lengths. Fig. 9(B) shows the compressed 3*3 microneedle array,
263 in which the microneedle tips are broken.

264 In Fig. 10(A), it can be seen that when the proline/silk fibroin ratio is 1/10, the displacement
265 load of the microneedle is the largest, which is 10.1 N; when the proline/silk fibroin ratio is 4/10,
266 the displacement load of the microneedle is the smallest, which is only 4.7 N.

267 In Fig. 10(B), when prolinamide/silk fibroin is 1/10, the maximum displacement load of the
268 microneedle is 11 N, and when 4/10, the minimum displacement load of the microneedle is 7 N.

269 In Fig. 10(C), when hydroxyproline/silk fibroin is 2/10, the maximum displacement load of the
270 microneedle is 13.3 N, and when it is 1/10, the minimum displacement load of the microneedle is
271 7.8 N.

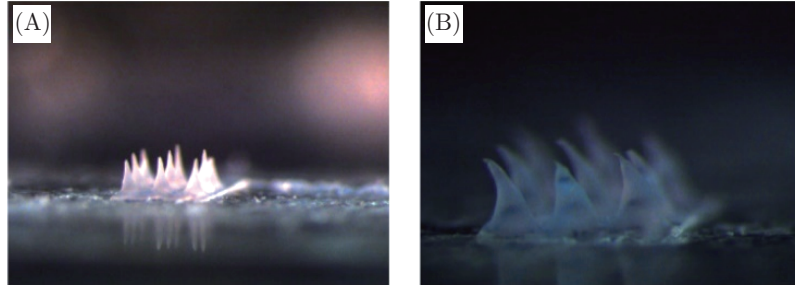


Fig. 9: Morphology of microneedle before and after mechanical test

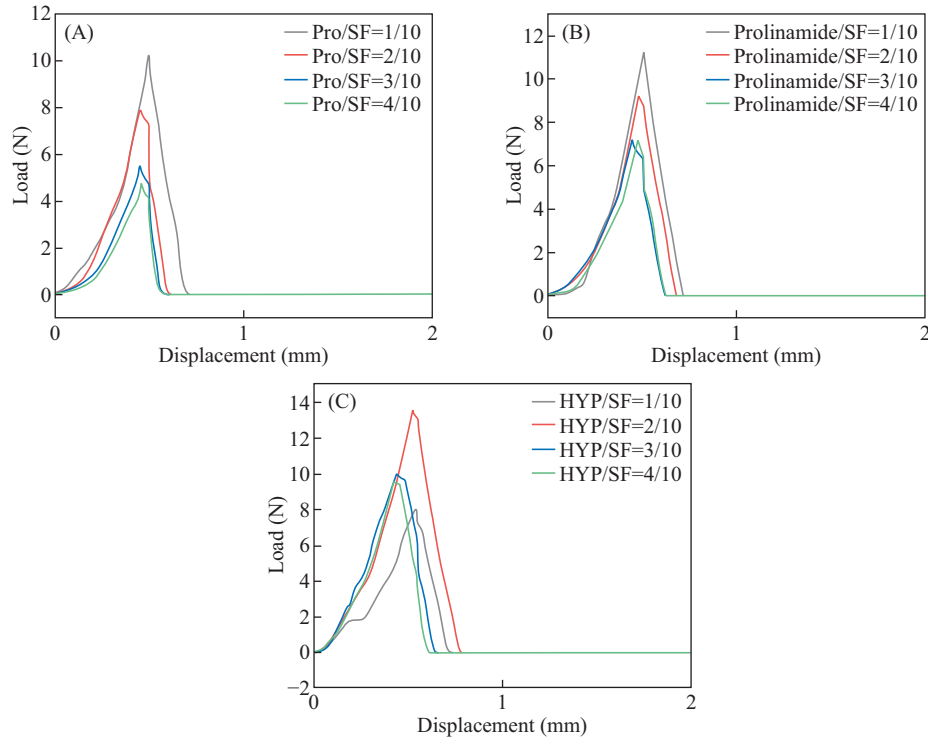


Fig. 10: Displacement load curve of proline and its derivative 3*3 microneedles: (A) proline/silk fibroin, (B) prolinamide/silk fibroin, (C) hydroxyproline/silk fibroin

272 Fig. 11 Significant difference analysis was performed using Tukey test. As can be seen from
 273 Fig. 11, there are significant differences among the three microneedles in any proportion. At
 274 2/10, the maximum compression strength of hydroxyproline/silk fibroin single needle reached 1.4
 275 N. According to the observation, except for 1/10, the compressive strength of single needle was
 276 hydroxyproline > prolinamide > proline. Therefore, the mechanical properties of hydroxyproline
 277 were the best, followed by prolinamide, and proline was the worst. In general, the breaking
 278 strength of microneedles decreased with the increase of amino acids, which was due to the fact
 279 that more small molecules were inserted between the molecular chains of silk fibroin to play the
 280 role of lubricant. When the external force acts on the microneedles, the relative slip of silk fibroin
 281 molecular chain is more likely to occur due to the action of small molecules, resulting in the
 282 decrease of its breaking strength. It has previously been reported that a single needle can break
 283 human skin with a force of 0.08 N. When the insertion force is 0.1-3 N, it is enough to penetrate
 284 the skin [17]. It can be seen from the figure, the strength of proline/silk fibroin microneedles,

285 hydroxyproline/silk fibroin microneedles and prolinamide/silk fibroin microneedles are very high,
 286 which is far greater than the force required to puncture the skin. Therefore, the microneedle
 287 has a good penetration performance, which can easily penetrate the cuticle of the skin without
 288 breaking, so as to achieve the role of drug delivery.

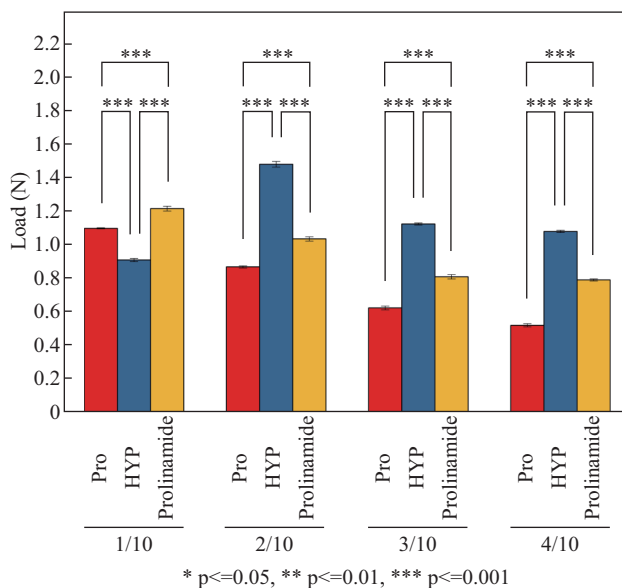


Fig. 11: Fracture strength of proline and its derivative/silk fibroin single microneedle

289 4 Conclusion

290 In this paper, silk fibroin was modified by proline and its derivatives to prepare swelling mi-
 291 croneedles. In the above experiments, it can be found that high proportion of hydroxyproline
 292 and silk fibroin will produce white crystals after blending, which is not suitable for micro needle
 293 materials. In the XRD and FTIR images, it can be found that silk fibroin blends with proline and
 294 its derivatives mainly show the Silk I crystal structure, so water molecules can enter the internal
 295 structure of the microneedle to produce swelling. In the swelling and dissolution diagram, when
 296 the proline/silk fibroin is 1/10, the swelling degree can reach 165%, but the dissolution rate is as
 297 high as 18%, so it is not suitable for swelling microneedles. When the mass ratio was 2/10, proline
 298 and its derivatives showed the same trend, with higher swelling degree and lower dissolution rate.
 299 In order to prepare high swelling microneedles, materials with low dissolution rate, high swelling
 300 degree and good mechanical properties need to be selected. When prolinamide /silk fibroin is
 301 2/10, the swelling degree is 150%, the dissolution rate is 5%, and the breaking strength of a single
 302 needle can reach 1 N. It can be used as a swelling microneedle for transdermal drug delivery.

303 References

- 304 [1] Abiandu I, Ita K. Transdermal delivery of potassium chloride with solid microneedles. *Journal of*
 305 *Drug Delivery Science and Technology*: 2019; 53: 101216-101223.

- 306 [2] Waghule T, Singhvi G, Dubey SK, et al. Microneedles: A smart approach and increasing potential
307 for transdermal drug delivery system. *Biomedicine & Pharmacotherapy*: 2019; 109: 1249-1258.
- 308 [3] Wiedersberg S, Guy RH. Transdermal drug delivery: 30+ years of war and still fighting. *Journal*
309 *of Controlled Release*: 2014; 190: 150-156.
- 310 [4] Prausnitz MR, Mitragotri S, Langer R. Current status and future potential of transdermal drug
311 delivery. *Nature Reviews Drug Discovery*: 2004; 3(2): 115-124.
- 312 [5] Lee H, Song C, Baik S, et al. Device-assisted transdermal drug delivery. *Advanced drug delivery*
313 *reviews*: 2018; 127: 35-45.
- 314 [6] Delgado-Charro MB, Guy RH. Effective use of transdermal drug delivery in children. *Advanced*
315 *Drug Delivery Reviews*: 2014; 73: 63-82.
- 316 [7] Rzhnevskiy AS, Singh TRR, Donnelly RF, et al. Microneedles as the technique of drug delivery
317 enhancement in diverse organs and tissues. *Journal of Controlled Release*: 2018; 270: 184-202.
- 318 [8] Henry S, McAllister DV, Allen MG, et al. Microfabricated Microneedles: A Novel Approach to
319 Transdermal Drug Delivery. *Journal of Pharmaceutical Sciences*: 1998; 87(8): 922-925.
- 320 [9] Ita K. Dissolving microneedles for transdermal drug delivery: Advances and challenges. *Biomedicine*
321 *& Pharmacotherapy*: 2017; 93: 1116-1127.
- 322 [10] Ita K. Ceramic microneedles and hollow microneedles for transdermal drug delivery: Two decades
323 of research. *Journal of Drug Delivery Science and Technology*: 2018; 44: 314-322.
- 324 [11] Donnelly RF, Singh TRR, Alkilani AZ, et al. Hydrogel-forming microneedle arrays exhibit antimi-
325 crobial properties: Potential for enhanced patient safety. *International Journal of Pharmaceutics*:
326 2013; 451(1): 76-91.
- 327 [12] Wu XL, Mao L, Qin DK, et al. Impact of Sterilization Methods on the Stability of Silk Fibroin
328 Solution. *Advanced Materials Research*: 2011; 311-313: 1755-1759.
- 329 [13] Altman GH, Horan RL, Lu HH, et al. Silk matrix for tissue engineered anterior cruciate ligaments.
330 *Biomaterials*: 2002; 23(20): 4131-4141.
- 331 [14] Li Y, Yuen CWM, Hu JY, et al. Analysis of the structural characteristics of nanoscale silk particles.
332 *Journal of Applied Polymer Science*: 2006; 100(1): 268-274.
- 333 [15] Yang H, Yang S, Kong J, et al. Obtaining information about protein secondary structures in
334 aqueous solution using Fourier transform IR spectroscopy. *Nature protocols*: 2015; 10(3): 382-396.
- 335 [16] Monti, P, Taddei, P, Freddi, G, et al. Raman spectroscopic characterization of Bombyx mori silk
336 fibroin: Raman spectrum of Silk I. Springer Netherlands: 2001; 32: 103-107.
- 337 [17] Davis SP, Landis BJ, Adams ZH, et al. Insertion of microneedles into skin: measurement and
338 prediction of insertion force and needle fracture force. *Journal of Biomechanics*, 2004, 37(8): 1155-
339 1163.